



## **SURFACE PLASMON RESONANCE IMMUNOSENSORS FOR ENVIRONMENTAL WATER ANALYSIS**

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## **SURFACE PLASMON RESONANCE IMMUNOSENSORS FOR ENVIRONMENTAL WATER ANALYSIS**



### **Outline presentation:**

- SPR
  - Introduction and Evanescent Field Sensors
  - Main features
  - Instruments
  - Functionalization on SPR sensors
- Pilot example of environmental analysis of organic pollutants
- Future and present work
- Conclusions

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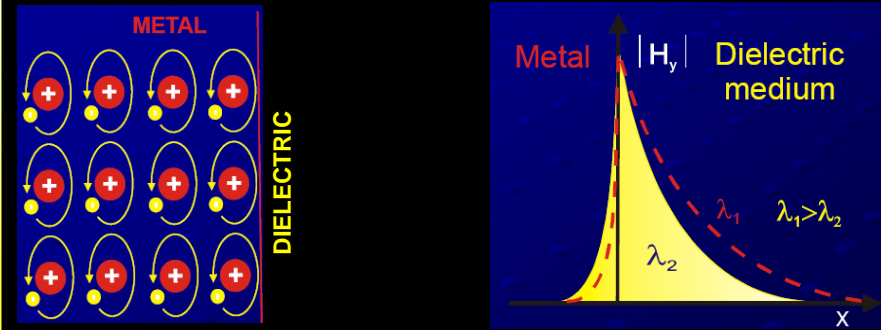


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### SPR theory

An incident monochromatic light at the interface between 2 substances with different refractive index can produce an evanescent wave



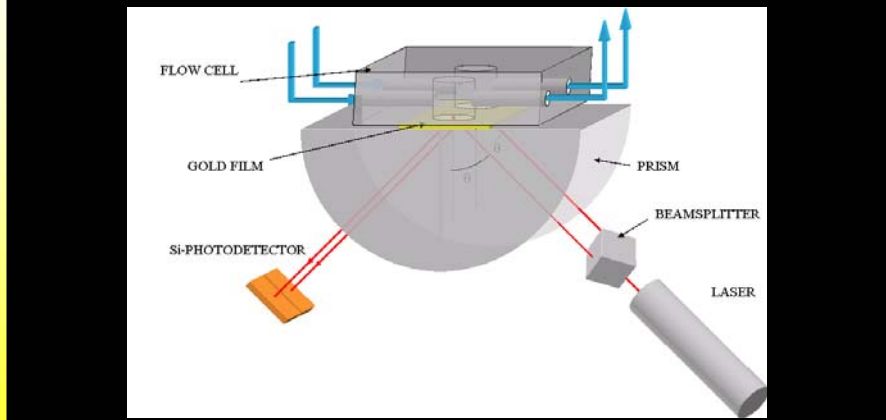
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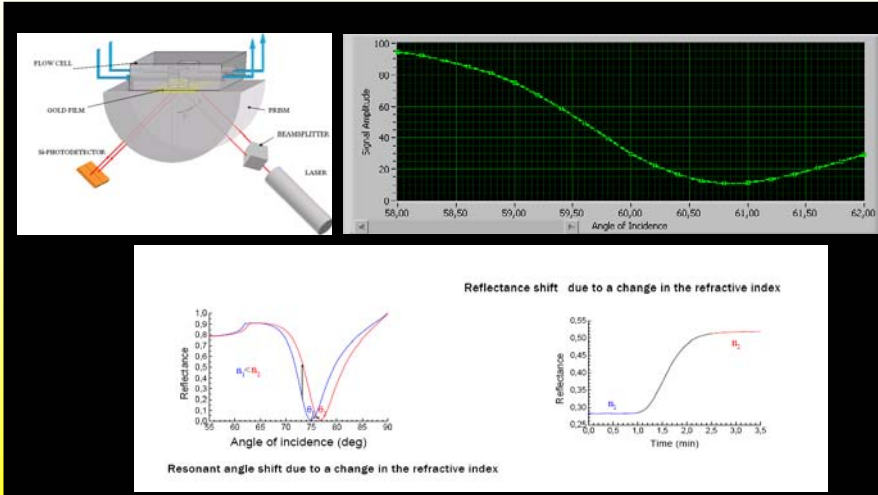
### SPR theory and evanescent field sensors



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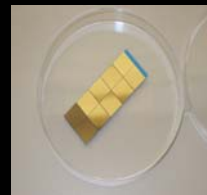
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### Biomolecular Recognition Elements in Evanescent Field Sensors:

SPR is a generic optical technology that can be combined with specific biological receptors against particular target analytes

- Antibodies → Immunosensor Chips
- Proteins
- DNA
- RNA
- MIPs (plastibodies)



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**Affinity SPR Biosensing: Detection Strategies**

**DIRECT: Medium size and large analytes**

**INDIRECT COMPETITIVE: Small analytes at low  
concentration**

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**Affinity SPR Biosensing: Detection Strategies**

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↓  
**Environmental analysis**

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**Surface Plasmon Resonance Biosensors:**

**Main Features**

- **Direct** (can detect analyte in one-step).
- **Real-time** (analyte's binding to sensor surface can be continuously monitored).
- **Label-free** (no fluorescent or radioactive labels are required for analyte's detection).
- **Minimum interaction length/volume required** (small sample volumes can be analyzed).
- **Generic technology** (combines generic optical technology with receptors *specific* against particular target analytes).

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**Laboratory SPR instruments based on angular  
Spectroscopy of surface plasmons:**

- **BIAcore (S51, 3000)**
- **Spreeta SPR sensor**
- **SENSIA**

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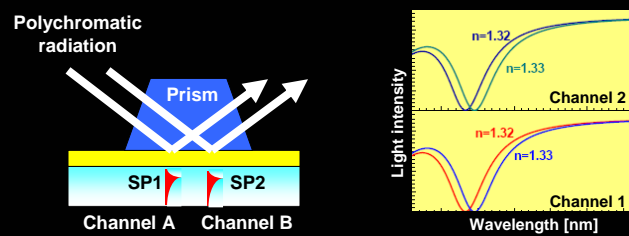
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**SPR Sensors with Referencing Channels:**



Multichannel SPR sensor with parallel sensing channels and  
SPR spectra observed

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### Functionalization of SPR Biosensors: Alkanethiol Attachment Chemistry

#### SAM formation

Mercaptoundecanoic acid 0.05 mM in EtOH

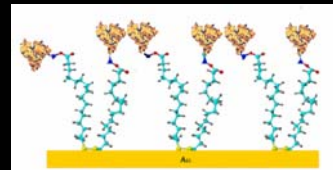
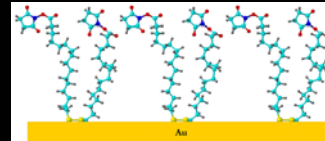
Formation N-Hydroxysuccinimide ester

- 1-Ethyl-3-(3-dimethyl-amino-propyl)carbodiimide hydrochloride (EDC) 0.2 M
- N-Hydroxysuccinimide (NHS) 0.05 M

#### Immunoreagent Immobilization

350+350  $\mu$ L of 10  $\mu$ g/L

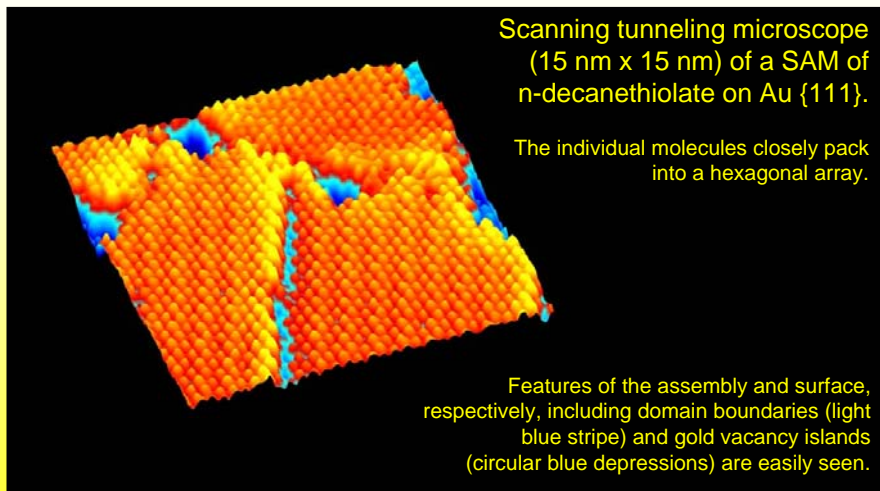
Blocking agent: Ethanolamine 1 M, pH 8.5



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Scanning tunneling microscope  
(15 nm x 15 nm) of a SAM of  
n-decanethiolate on Au {111}.

The individual molecules closely pack  
into a hexagonal array.

Features of the assembly and surface,  
respectively, including domain boundaries (light  
blue stripe) and gold vacancy islands  
(circular blue depressions) are easily seen.

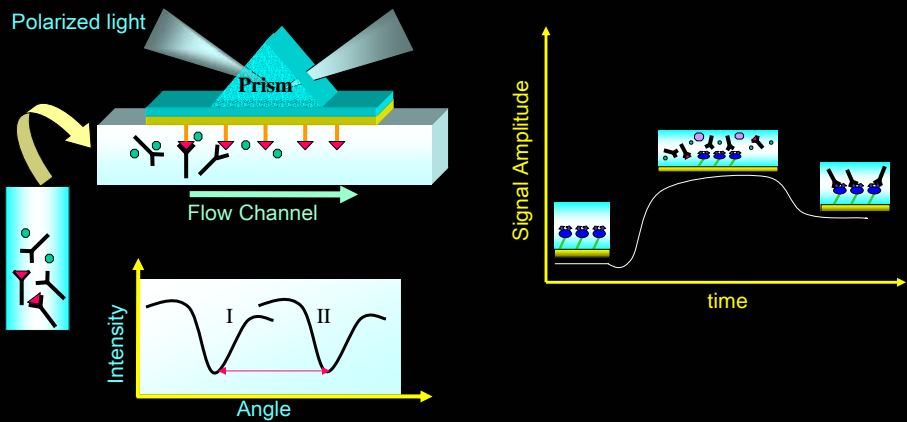
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### Indirect immunoassay



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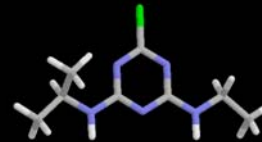
### PILOT ANALYTE: ATRAZINE

#### Development of immunosensor chips

- Alkanethiol Attachment for SAM formation
- 2d-BSA atrazine conjugate

#### Immunoassay optimization

- Indirect format assay
- Purified polyclonal antibodies against Atrazine



#### Validation

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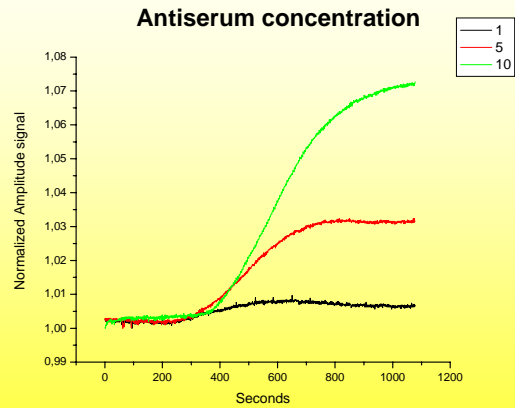


## SURFACE PLASMON RESONANCE IMMUNOSENSORS FOR ENVIRONMENTAL WATER ANALYSIS



### Immunoassay optimization:

- Antiserum concentration
- Incubation time
- Elution
- Buffers
- Regeneration



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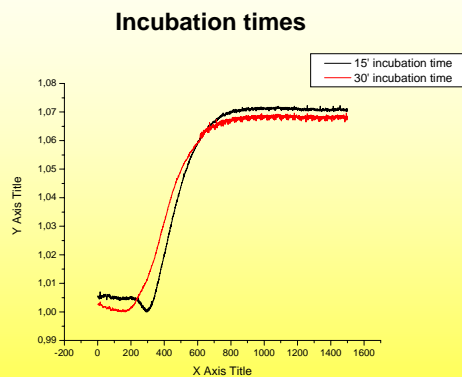


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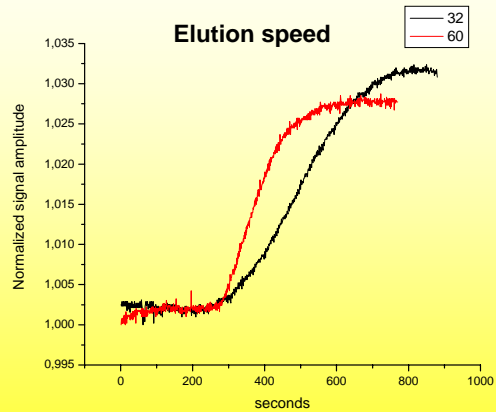


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### Immunoassay optimization:

- Antiserum concentration
- Incubation time
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### Regeneration:

- HCl 0.05 M
- HCl 0.75 M
- HCl 0.1 M
- HCl 0.2 M
- NaOH 0.05 M
- NaOH 0.1 M

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### Regeneration:

- HCl 0.05 M
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- HCl 0.2 M
- NaOH 0.05 M
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\* More than 120 regeneration cycles can be performed

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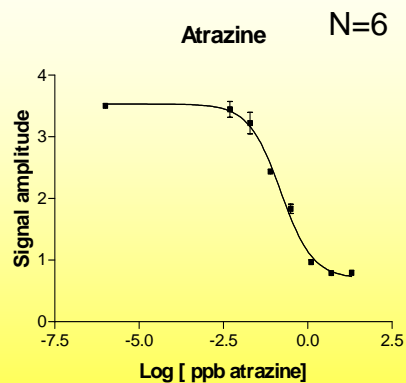


### Optimized Assay

Acetic-acetate  
Assay buffer: PBS  
Coating: 10  $\mu\text{g/L}$  2d-BSA  
Ab: 10  $\mu\text{g/L}$   
Incubation time 15'  
Elution speed 32 (23  $\mu\text{L/min}$ )  
Regeneration: NaOH 0.1 M

**IC<sub>50</sub> = 0.17**

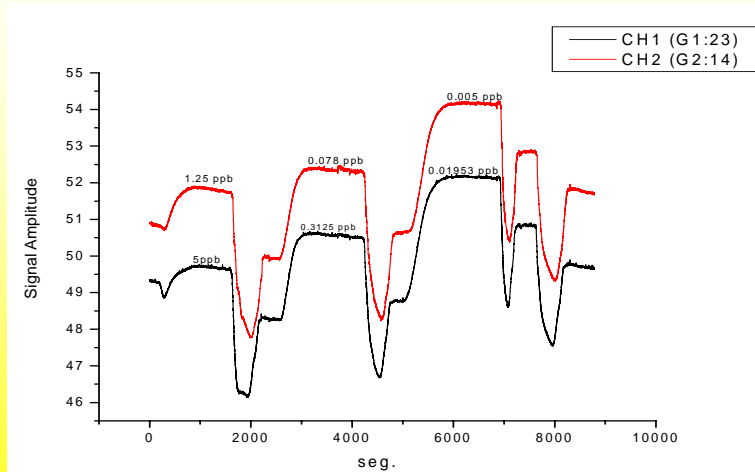
**LOD = 0.02 ppb = 20 ng/L**



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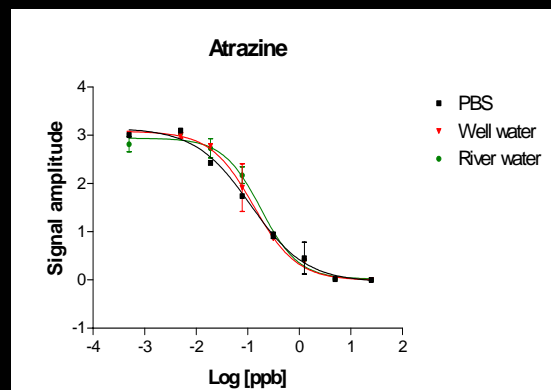
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#### Matrix effects study



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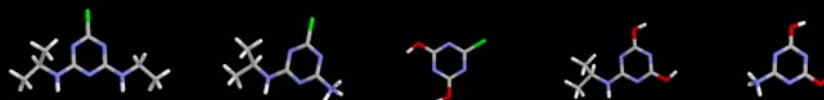


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**Cross Reactivity Studies**

Compound	CR(%) = [IC50/IC50(atrazine)] 100
Atrazine	100
Simazine	25
Terbutylazine	23
Desethyl-atrazine	5



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**VERIFICATION:**

Sample	SPR µg/L	SPE- HRGC/MS µg/L	Sum of Triazine µg/L	Sample	SPR µg/L	SPE- HRGC/MS µg/L	Sum of Triazine µg/L
1	0.05	0.05	0.25	8	0.10	0.07	0.29
2	0.10	0.08	0.13	9	0.12	0.07	0.27
3	1.00	0.82	1.07	10	0.06	0.05	0.1
4	0.26	0.24	0.46	11	0.10	0.07	0.15
5	0.22	0.19	0.51	12	0.07	0.06	0.09
6	0.20	0.20	0.59	13	0.06	0.06	0.09
7	0.11	0.07	0.27	14	0.10	0.07	0.11

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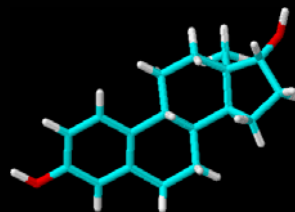
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**Future Work:**

**Development of immunosensing chips for analysis of emergent pollutants.**

- Monoclonal antibodies
- Estrogens: E<sub>2</sub>, E<sub>1</sub>, EE<sub>2</sub>
- Pharmaceutical compounds



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**Conclusions:**

- **Optical biosensors based on spectroscopy of surface plasmons provide label-free, fast, specific and sensitive measurements and it is a complementary approach to traditional laboratory analytical techniques.**
- **Real time analysis**
- **Inexpensive**
- **Portable systems**
- **Various immobilization procedures can be performed to meet needs of specific applications.**

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**Conclusions:**

- A rapid method for triazine screening have been developed
- This method is high sensitive LOD 20 ng/L
- No sample treatment is required just filtration and buffer adjustment
- Good agreement was observed validated vs. HRGC-MS

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This work has been supported by the EU project SWIFT-WFD, contract SSPI-CT-2003-502492.

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SENSIA

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**THANK YOU  
VERY MUCH !!!!!**

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